

Introduction

The ARC-sparX Platform and ACLX001*

The ARC-sparX platform separates the antigen-recognition and killing functions of conventional CAR-T therapy and is comprised of two key components:

- **1.** SPRX001 (soluble protein antigen-receptor X-linker) protein: binds BCMA on diseased cells and flags those cells for destruction.
- 2. ARC-T Cells (Antigen Receptor Complex T Cells): bind and kill SPRX001 flagged cells. **3.** ACLX001 is the combination of the ARC-T cells and SPRX001.



SPRX001 protein

- No inherent therapeutic activity
- Contains two BCMA-targeting domains
- Binds BCMA+ multiple myeloma cells Engineered to minimize immunogenic potential
- "TAG" is a fragment of human alpha fetoprotein (AFP)

ARC-T cell

- CAR is comprised of the extracellular novel binding domain fused to CD8α spacer/ transmembrane region fused to 4-1BB/ CD3ζ signaling domains
- Only activated upon formation of a tricomplex of ARC-T + SPRX001 + BCMA positive multiple myeloma cells
- Same viral vector regardless of antigen target

Results

ACLX001 demonstrates dose- and target-dependent cytokine production, degranulation, cytotoxic activity, and proliferation



sequentially incubated with SPRX001, BCMA-His-AviTag, then streptavidin-PE to detect binding of BCMA. An anti-FLAG antibody was used in parallel to detect ARC expression.

B. SPRX001 specifically binds BCMA-expressing cells. NCI-H929 cells (wild-type or BCMA-CRISPR-KO) were directly stained with AlexaFluor647 labeled SPRX001.

C. The trimolecular complex of BCMA-SPRX001-ARC induces a cytokine and degranulation response from ARC-T cells. ARC-T cells were cultured with increasing concentrations of SPRX001, with anti-CD107A antibody and monensin for 5 hours in the presence or absence of H929-WT or H929-BCMA^{-/-}. Cells were then subject to fixation/intracellular staining for IL-2, IFN-y, and TNF-α.

ACLX-001*, a novel BCMA-targeted CAR-T cell therapy that can be activated and silenced in vivo with soluble protein adapters in a dose dependent manner

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D. The trimolecular complex of BCMA-SPRX001-ARC induces a cytotoxic response from ARC-T cells. ARC-T cells were cultured overnight with a dose titration of SPRX001 (or the negative control SPRX-α3D) and H929-WT, H929-BCMA^{-/-,} or NALM6-BCMA (all GFP and luciferase expressing). After culture, cytotoxicity was assessed based on remaining luciferase activity in cell pellet.

E. The trimolecular complex of BCMA-SPRX001-ARC drives ARC-T cells proliferation in an SPRX001 dose dependent manner. ARC-T cells were rested overnight in IL-2 free media, then stained with violet proliferation dye to assess cell division. Dye-stained ARC-T cells were cultured for 96-hours with a dose titration of SPRX001 (or the negative control SPRX-α3D) and NALM6-BCMA or SPRX001 and T cells alone. After 96-hours, cells were stained for CD3 and total T cells (CD3+GFP-) were enumerated from individual wells of a 96-well plate using a high throughput sampler.

ACLX001 eliminates MM.1S tumor cells in vivo comparable to a **BCMA-targeted CAR-T**



later (Day 0), mice received 5×10⁶ ddBCMA-CAR cells (group 1) or 5×10⁶ ARC-T cells (groups 2-5). Mice that received ARC-T cells were dosed daily with sparX proteins IP at indicated dose levels for 21 days. Animals were subsequently imaged by IVIS throughout. **B.** Total tumor burden was quantitated from the images in (A).

ACLX001 exhibits comparable activity in vivo with intermittent dosing of SPRX001 in a disseminated model of BCMA+ B-cell leukemia (NALM6-BCMA)



A. ACLX001 causes regression of NALM6-BCMA tumors in vivo with intermittent dosing of SPRX001. NSG mice were engrafted with 1×10⁶ NALM6-BCMA cells expressing GFP and Luciferase. 10 days later (Day 0), mice were engrafted with 5×10⁶ ARC-T cells and were dosed with the negative control SPRX-α3D daily or with the SPRX001 daily, every other day, twice per week, or weekly for 21 days. Animals were imaged on IVIS throughout.

B. Total tumor burden was quantitated from the images in (A). Grey shading indicates the period of sparX-protein administration.

C. The in vivo residence time of SPRX001 on NALM6-BCMA tumor cells was assessed in mice following I.V. SPRX001 administration. Femurs were collected at various timepoints post-injection, flushed, samples fixed, and SPRX001-binding was detected on tumor cells via an anti-tag antibody. Binding is presented as percent positive (left). Representative histograms for each time point are also presented (right).

ARC-T cell kinetics and memory cell differentiation vary widely in vivo



A. ACLX001 (ARC-T + SPRX001) halts expansion and causes regression of NALM6-BCMA in vivo. NSG mice (NOD-scid IL2Rg^{null}) were engrafted with 1×10⁶ NALM6-BCMA cells expressing GFP and Luciferase. 10 days later (Day 0), animals were imaged on IVIS and randomized. Mice were engrafted with ARC-T cells (T-cell receptor wild type or CRISPR-KO) or Mock transduced T cells (T-cell receptor wild type or CRISPR-KO). All mice were dosed with SPRX001 every other day (q.o.d.) for 28 days. Animals were imaged by IVIS throughout.

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- **B. (Left)** Absolute number of circulating GFP+ tumor cells per µl of whole blood were monitored throughout the course of the experiment. (**Right**) Total tumor burden was quantitated from the dorsal images in (A).
- **C.** Absolute number of circulating total T cells (left) or ARC+ T cells (right) per µL of whole blood were monitored throughout the course of the experiment. After day 14 post-T-cell transfer, T-cell expansion is driven primarily through GvHD in the TCR-WT groups in this experiment.
- **D.** T cells were assessed for their expression of CD45RA and CD62L to determine their phenotype prior to transfer. CD8 T cells were primarily of the naïve/stem memory phenotype, while CD4 T cells were a mixture of naïve/stem memory and central memory.
- **E.** SPRX001 drives effector and memory phenotype differentiation of ARC-T cells as they are activated and expand in vivo. T cells were monitored for their differentiation into effector and memory phenotype cells seven days post-transfer based on their expression of CD45RA and CD62L.
- **F.** Effector and memory phenotype differentiation of ARC-T or Mock transduced (TCR-WT or TCR-KO) were tracked throughout the course of experiment. As in **(D)**, T cells were monitored for their differentiation into effector and memory phenotype based on their expression of CD45RA and CD62L throughout the course of the experiment. CD8+ T cells are presented on the top and CD4+ T cells are presented on the bottom. After day 14 post-T-cell transfer, T-cell differentiation is driven primarily through GvHD in the TCR-WT groups as compared to the TCR-KO groups.

Conclusions

- ACLX001 is a novel cell therapy for BCMA+ malignancies
- The SPRX001 protein imparts BCMA-binding and -targeting capacity to ARC-T Cells
- Only when SPRX001 bridges ARC-T cells to BCMA on target cells are ARC-T cells activated to produce cytokines, have cytotoxic activity against BCMA+ targets, or proliferate in response to BCMA+ target cells.
- ACLX001 eliminates MM.1S tumor cells in vivo comparable to a BCMA-targeted CAR-T
- SPRX001 dosed daily, every other day, or twice per week eliminated measurable NALM6-BCMA tumor burden
- Despite rapid reduction in receptor occupancy on the BCMA+ tumor cells, intermittent dosing still achieves tumor clearance
- ARC-T cell kinetics and memory cell differentiation vary widely in vivo when driven by different stimuli

*The data in this poster were generated with related but distinct reagents from the ACLX001 clinical reagents and are meant to support the clinical development of this product.